

Anti-HHV-6 Antibodies in Normal Population and in Cancer Patients in India

K.R. SHANAVAS,¹ V. KALA,¹ D.M. VASUDEVAN,¹ T. VIJAYAKUMAR,²
and M. YADAV³

¹*Department of Biochemistry¹, Medical College, Trichur, Kerala, India*

²*Department of Science, Technology and Environment, Trivandrum, Kerala, India*

³*Department of Genetics and Cellular Biology, University of Malaya, 59100 Kuala Lumpur, Malaysia*

ABSTRACT - The prevalence and titre of IgG antibodies to human herpesvirus type 6 (HHV-6) were assayed in the serum samples from normal subjects and patients with Hodgkin's lymphoma (HL), non-Hodgkin's lymphoma (NHL), acute lymphoblastic leukaemia (ALL) and oral cancer (OC) using immunofluorescence and immunoperoxidase techniques. This forms the first study on the sero-prevalence and titre of antibodies to HHV-6 in India. There was no considerable difference in the prevalence (76%) and titre (10-160) of the antibodies in normal population from those reported for normal adults in other parts of the world. All the HL and ALL patients studied showed no significant elevation in the antibody titre, though a slight increase in the prevalence (95%) was noted. Antibody titre and prevalence were found highly elevated in OC. OC remained totally unstudied for the presence of anti-HHV-6 antibodies, and this is the first report of elevated levels of the antibody in this cancer. The role of HHV-6, if any, in the pathogenesis of OC is worth investigating.

INTRODUCTION

SALAHUDDIN et al. (1986) isolated a new virus from peripheral blood lymphocytes of 4 patients with lymphoproliferative disorders and 2 patients with AIDS. The virus was named Human B-Lymphotropic Virus (HBLV). In some of the patients from whom the virus was derived, there were B-cell lesions. Subsequent molecular biological and structural studies, however, revealed that this virus has the typical features of a Herpes virus which predominantly infects T-lymphocytes. The virus was, therefore, renamed as human herpesvirus type 6. Seroepidemiological studies have shown HHV-6 to be ubiquitous in human populations, and it has been isolated from the blood and saliva of normal adults (Harnett 1990; Levy et al. 1990; Yadav, 1991).

The prevalence of the antibodies to HHV-6 in normal human populations worldwide ranges from 60 - 85% (Yadav 1991). The antibody titre ranges from 10 - 360, and may decrease in the individuals above 40 years of age. Elevated levels of anti-HHV-6 antibodies have been found in patients with Exanthem Subitum, chronic fatigue syndrome, Kikuchi lymphadenitis, a type of infectious mononucleosis, AIDS, lymphoid malignancies and in organ transplant patients receiving immunosuppressive drug therapy (Ablashi et al. 1988).

Studies have been made on the seroprevalence and titre of the antibody to HHV-6 in normal subjects as well as in cancer patients (Saxinger et al. 1988; Okino et al. 1988; Clark et al. 1990, b; Yadav, 1991). Yadav (1991) reported not only a high prevalence rate of anti-HHV-6 antibodies in lymphomas but also high titres of 160 to >512.

against a low titre of 10 to 320 in normal adults. Studies have not yet been made in India either on the seroprevalence or on the titre of anti-HHV-6 antibodies in normal population or in cancer patients. Hence this preliminary study.

MATERIALS AND METHODS

Twenty-eight patients with Hodgkin's lymphoma (HL), 59 non-Hodgkin's lymphoma (NHL), 51 acute lymphoblastic leukaemia (ALL) and 127 patients with oral cancer (OC) were included in the study. Seventy five age-matched healthy subjects were also included in the study as control. None of the patients had undergone any type of treatment for cancer prior to the blood collection for the study. Venous blood was collected, serum separated, inactivated at 56°C for 30 min, and as far as possible, the tests were carried out on the day of collection itself; or else the serum was stored at -70°C for a maximum period of 2 weeks.

The presence of IgG antibodies to the viral capsid antigen of the Z29 strain of HHV-6 and its titre in serum were evaluated using indirect immunofluorescence assay and immunoperoxidase techniques. Molt-3 cells, infected with HHV-6 in culture, were used as the source of HHV-6 viral capsid antigen (Yadav et al. 1990a). The Molt-3 cells, maintained in RPMI-1640 medium with 10% fetal calf serum (5% CO₂ at 37°C), were infected with Z29 strain OF HVV-6 in the presence of polybrene (2 g/ml). Post-infectionally, the Molt-3 cells were grown in the medium supplemented with 5% fetal calf serum until 50% or more atypical blast cells were present in the culture. The Molt-3 cells were harvested,

washed 3 times in phosphate buffered saline (PBS), on teflon-coated multi-well slides, fixed in cold for 10 min and ~~stored~~ stored at -20°C until used.

The indirect immunofluorescence assay was done following Yadav and Ablashi (1990). Starting from 1/10, a series of 2-fold dilutions of the sera was prepared. The slides with the smears of HHV-6 infected Molt-3 cells were incubated with the diluted sera at 37°C for 30 min in a humid chamber, washed with PBS, then incubated with fluorescein conjugated anti-human IgG antibodies at 37°C for 30 min, washed thoroughly in PBS, dried, mounted with fluorescent mounting medium (Pan-data Systems, USA) and coverslip slides were observed under the 100x objective fluorescence microscope. Anti-HHV-6 antibody positive and negative control sera were incorporated in every batch.

Immunoperoxidase staining was carried out following the method of Gregory (1988). Endogenous peroxidase activity was blocked by treating the smears with 0.03% H_2O_2 in methanol for 10 minutes. Incubation with the test sera was done as detailed above. The smears were washed, and then incubated with horse radish peroxidase conjugated anti-human IgG antibodies at 37°C for 45 min and washed thoroughly in PBS. Subsequently, PBS containing 0.05% diaminobenzidine tetrahydrochloride and 0.01% hydrogen peroxide, was added as the substrate and incubated at room temperature for 5 min and washed thoroughly in PBS. The slides were mounted with aqueous mounting medium (BDH), covered with coverslips and studied under the objective of a microscope.

RESULTS

The results of the study are presented in Table 1 and

Subjects	Prevalance	Antibody titre	
		Range	Mean
Normal Control (n = 75)	76%	10-160	47
NHL (n = 59)	95%	10-320	88
HL (n = 51)	100%	320-5120	1109
ALL (n = 51)	100%	160-5120	1208
OC (n = 127)	96%	80-10240	2042

Table 1 : Antibody prevalance and titre to human herpesvirus-6 (HHV-6) in the sera of normal subjects and of cancer patients.

NHL : Non Hodgkin's lymphoma; HL: Hodgkin's lymphoma; ALL: acute lymphoblastic leukaemia; OC: oral cancer

Subjects	Total Numbers	No. of persons having the antibody titre of				
		<10	10-80	160-640	1280-5120	10240
Normal	75	18	37	10	-	-
NHL	59	3	36	20	-	-
HL	28	-	-	12	16	-
ALL	51	-	-	35	16	-
OC	127	4	7	51	54	11

Table : 2. Anti-HHV-6 antibody titres in the sera of normal subjects and of patients

NHL : Non-Hodgkin's lymphoma; HL: Hodgkin's lymphoma; ALL:Acute lymphoblastic leukaemia;

OC : Oral cancer

The results obtained by immunofluorescence and immunoperoxidase techniques were identical in the initial standardisation studies, and hence the results are not given separately. The prevalence of anti-HHV-6 antibodies in normal control was 76%, and the titre ranged from 10 to 160 (mean, 47). In HL and ALL, the antibodies could be detected in all the serum samples, with titres of 320 to 5120 (mean, 1109) and 160 to 5120 (mean, 1208) respectively. The prevalence of the antibody in the sera of NHL and OC patients was 95% and 96% respectively. While the antibody titre was relatively low in NHL (10 to 320; mean, 88); it was seen to be very high in OC (80 to 10250; mean, 2042).

DISCUSSION

Yadav (1991) has reported a high prevalence (80%) of anti-HHV-6 antibodies in the sera of HL and ALL patients. In the present study, we observed the presence of the anti-viral antibody in all the patients with HL and ALL. The titre of the antibody observed in the present series of patients was same in HL and ALL, but was considerably higher than the titres reported by Yadav (1991). It is known that in HL and ALL, the blast cells include those with T-cell markers (Devita, 1973; Kadin, 1985; Nadkarni et al. 1978). HHV-6 has been reported to infect T-lymphocytes at various stages of differentiation in vivo (Yadav, 1991). The high prevalence and titre of anti-HHV-6 antibody in the sera of HL and ALL patients may be due to an increase in the number of cells with HHV-6 receptors in these diseases.

Oral cancer is the most prevalent carcinoma in India, and forms about 27% of the total cancer incidence in this part of the country (Padmanabhan and Vasudevan, 1981). There are

no previous reports on the association of HHV-6 with cancer. In the present study, anti-HHV-6 antibody observed in 96% of the serum samples from OC patients. The titre of the antibody was highly elevated in the majority of the patients. Saranath et al (1985) Vijayakumar and Vasudevan (1985) have observed derangement of cell-mediated immunity in OC patients. This may be due to the reactivation of the virus which in turn could result in the elevated levels of the antibody. HHV-6 has been reported to be cytopathic to the host T-cells, which are important in cell-mediated immunity. A reactivated infection, hence, could possibly increase the severity of the disease by further weakening the already impaired immune response. Or the derangement in immunity may be due to the newly acquired HHV-6 infection. However, the exact role of HHV-6 in OC remains to be determined through further investigations.

ACKNOWLEDGEMENT- Our grateful thanks to Dr. D.V. Alexander, National Cancer Institute, USA, for advice and to Mr. Anil Kumar, Department of Biochemistry, Medical College, Trichur, for technical assistance.

REFERENCES

1. Ablashi DV, Joseph SF, Buchbinder A, Hellmuth G, Nakamura S, Llana T, Lusso P, Kaplan M, Dahlborg G, Memon S, Imam E, Ablashi KL, Markham PD, Kramarsky R, Krueger GRE, Biberfeld P, Wong-stall F, Sala-i-Martin X, Gallo RC. Human B-lymphotropic virus (human herpesvirus-6). J Virol Methods 21, 29-48, 1988.
2. Clark DA, Alexander FE, Mckenney PA, Robert

O'Brien C, Jarret RF, Cartwright RA, Onions DE. The seroepidemiology of human herpesvirus-6 (HHV-6) from a case-control study of leukaemia and lymphoma. *Int J Cancer* 45, 829-833, 1990.

3. De Vita VT. Lymphocyte reactivity in Hodgkin's disease: a lymphocyte civil war. *New Engl J Med* 289, 801-802, 1973.
4. Harnett GB, Farr J, Pietroboni GR, Bucens MR. Frequent shedding of human herpesvirus-6 in saliva. *J Med Virol* 30,128-130, 1990.
5. Jones FL, Gregory J. Immunoperoxidase methods. In: Catty D, Ed-'Antibodies, a practical approach, Vol.II', New York IRL Press, 155-178, 1988.
6. Kadin ME. Common activated helper T-cell origin for lymphomatoid papulosis, mycosis fungoides and some types of Hodgkin's disease. *Lancet* ii, 804-865, 1985.
7. Levy JA, Ferro F, Greenspan D, Lennette ET. Frequent isolation of HHV-6 from saliva and high seroprevalence of the virus in population. *Lancet* i, 1047-1050,1990.
8. Nadkarni JJ, Nadkarni JS, Advani SH. Surface markers in human lymphoid leukaemias. *Indian J Cancer* 15, 43-47,1978.
9. Okuno T, Takahashi K, Balachandra K, Shiraki K, Yamanishi K, Takahashi M, Baba K. Seroepidemiology of human herpesvirus-6 in normal children and adults. *J Clin Microbiol* 27, 651-653, 1989.
10. Padmanabhan TK, Vasudevan DM. A statistical analysis

of cancers registered at the Regional Cancer Centre, Trivandrum. Indian J cancer 18, 189-196, 1982.

11. Salahuddin ZS, Ablashi DV, Markham PD, Josephs Sturzenegger S, Kaplan M, Halligan G, Biberfeldt Wong-Staal F, Kramarsky B, Gallo RC. Isolation of a new virus HBLV in patients with lymphoproliferative disorders. Science 234, 596-601, 1986.
12. Saranath D, Mukhopadhyaya R, Rao RS, Fakhri AR, Sankar SL, Gangal SG. Cell-mediated immune status in patients with squamous cell carcinoma of the oral cavity. Cancer 56, 1062-1070, 1985.
13. Saxinger C, Polesky H, Eby N, Grufferman S, Meltzer R, Tegtmeyer G, Parekh V, Memon S, Hung G. Antibody reactivity with HBLV (HHV-6) in U.S. population. Virol Methods 21, 199-208, 1988.
14. Vijayakumar T, Vasudevan DM. High affinity rose bengal forming cells in carcinoma of the oral cavity, uterine cervix and breast. Cancer letters 27, 339-345, 1985.
15. Yadav M. Active human herpesvirus-6 infections associated diseases. Mal J Child Health 3, (in press) 1991.
16. Yadav M, Ablashi DV. Nasopharyngeal carcinoma: presence of elevated antibody to human herpesvirus-6 in patients. Asean J Clin sci 10, 81-84, 1990.
17. Yadav M, Ponniah K, Umamaheswari S. IgG antibody to human herpesvirus-6 in Malaysians. J. Biosci 1, 59-63, 1990a.

18. Yadav M, Umamaheswari S, Ablashi DV. Low prevalence of antibody to human herpesvirus-6 (HHV-6) in Kadazans. Southeast Asian J Trop Med Hlth 21, 259-263, 1990b.

Correspondence and reprint requests to :

Dr. D.M. Vasudevan

Professor and Head

Department of Biochemistry

Medical College

Trichur - 680 596

Kerala, India